

rGH [¹²⁵I] RIA KIT

(REF: RK-551)

For Research Use Only. Not for use in diagnostic procedures.

The [¹²⁵I]rGH RIA system provides direct quantitative *in vitro* determination of rat Growth Hormone in plasma, tissue and cell culture. rGH can be assayed in the range of 1.6-100 ng/ml. Each kit contains materials sufficient for 120 determinations permitting the construction of one standard curve and the assay of 50 unknowns in duplicate.

Introduction

Rat growth hormone (rGH) (also called somatotrophin), is a pituitary hormone whose molecular weight is approximately 22 000. It is a simple polypeptide hormone with no carbohydrate or lipid moieties, whose amino acids sequence is species specific.

As its name implies, this hormone stimulates growth and in terms of weight GH is the major hormone of the anterior pituitary. Unlike most of the pituitary hormones, GH does not have a single target gland, instead it can influence a variety of tissues. It promotes growth of bone, soft tissue and viscera. This is particularly important in young, growing animals, though GH seems to be relatively unimportant in the foetus and in the neonate. GH has also been shown to play an auxiliary role in lactation in rats, which may be important in times of prolactin insufficiency.

There are two principle hypothalamic factors controlling GH release, somatostatin and GH releasing factor (GRF). The ability of the pituitary gland to release GH in response to GRF is dependent on the amount of somatostatin present. In addition to hypothalamic peptides controlling GH release, there are a number of other peripheral factors affecting GH secretion (for example gonadal or adrenal steroids, thyroid hormone or insulin), as well as the possibility of a direct feedback of GH itself on its own release.

In the rat, males and females have very different patterns of GH secretion. The male rat has an intermittent pattern of GH release with large amplitude pulses occurring regularly at 3-hourly intervals throughout the day and night. In contrast, the female rat has much more frequent and smaller pulses.

Principle of method

This assay is based on the competition between unlabelled rGH and a fixed quantity of [¹²⁵I] -labelled rGH for a limited number of binding sites on a specific antibody. With fixed amounts of antibody and radioactive ligand, the amount of radioactive ligand bound will be inversely proportional to the concentration of added non-radioactive ligand.

The antibody bound rGH is then reacted with the separating second antibody reagent. Separation of the antibody bound fraction is effected by either magnetic separation or centrifugation of the separating reagent suspension and decantation of the supernatant.

Measurement of the radioactivity in the pellet enables the amount of labelled rGH in the bound fraction to be calculated. The concentration of unlabelled rGH in the sample is then determined by interpolation from a standard curve. The standard curve and samples should be prepared simultaneously.

Contents of the kit

1. 1 vial TRACER, lyophilized, reconstitution with 12.5 ml assay buffer, containing ~ 76 kBq, 2.05 μCi [¹²⁵I]rGH. The final solution contains [¹²⁵I]rGH in 0.025 M phosphate buffer pH7.5 containing 0.1%(w/v) sodium azide. Store at 2-8 °C.

2. 1 vial STANDARD, lyophilized, reconstitution with assay buffer, volume stated on the vial label. The final solution contains rGH at a concentration of 100 ng/ml. Store at 2-8 °C.

3. 1 vial ANTISERUM, lyophilized, reconstitution with 12.5 ml assay buffer. The final solution contains rabbit anti-rGH serum in 0.025 M phosphate buffer pH7.5 containing 0.1%(w/v) sodium azide. Store at 2-8 °C.

4. 1 vial ASSAY BUFFER concentrate (10.0 ml), dilution to 100 ml. On dilution this will give 0.025 M phosphate buffer, pH7.5, containing 0.1%(w/v) sodium azide. Store at 2-8 °C.

5. 1 vial SEPARATING SECOND ANTIBODY REAGENT (58 ml), ready for use, containing donkey anti-rabbit serum coated on to magnetisable polymer particles with sodium azide, colour-coded, blue-green. Store at 2-8 °C.

Pack leaflet

Materials, tools and equipment required

Pipettes or pipetting equipment with disposable tips (100μl, 200μl, 400μl, 500μl, 2.0ml and 12.5ml); disposable polypropylene or polystyrene tubes (12 x 75 mm); refrigerator; glass measuring cylinder (100ml); test tube rack; vortex mixer; plastic foil; separators, comprising magnetic base and assay rack - these are for use in the magnetic separation protocol; adsorbent tissue; gamma counter.

Note: For the centrifugal protocol, the following additional equipment will be required:

Decantation racks; refrigerated centrifuge capable of >1500 xg.

Specimen collection, storage

This section is provided for guidance only. It remains the investigator's responsibility to validate the chosen sample collection technique.

Blood samples

It is advised that if measurements are to be made on plasma samples, blood should be collected into tubes containing heparin (<20 units per ml of blood). Blood should be centrifuged immediately to remove cells and the plasma stored below -15°C prior to analysis. Serum samples can also be assayed with this kit. Samples may need to be diluted prior to assay depending on the expected concentration. Assay buffer remaining after reconstitution of assay components may be used for this.

Tissue samples

Pituitary glands should be frozen immediately after removal and stored at -15°C to -30°C until required. Prior to analysis, glands should be homogenised or sonicated in phosphate buffered saline solution. Samples may need to

be diluted depending on the expected concentration. Assay buffer remaining after reconstitution of assay components may be used for this.

The efficiency of various extraction procedures for the estimation of rat anterior pituitary hormone content has recently been extensively examined.

Cell cultures

Cell culture media should be collected and stored below -15°C prior to analysis. Samples may need to be diluted depending on the expected concentration. Assay buffer remaining after reconstitution of assay components may be used for this.

Preparation of reagents, storage

Storage: see *Contents of the kit*. At these temperatures each reagent is stable until expiry date. The actual expiry date is given on the package label and on the quality certificate.

Preparation: Equilibrate all reagents and samples to room temperature prior to use. Assay buffer: Warm the bottle containing assay buffer concentrate to 40°C or until the gel-like material melts. Temperatures above 60°C should be avoided. Transfer the contents of the bottle, with washings, to a 100 ml measuring cylinder and dilute to 100 ml with distilled or deionised water. Mix well. Assay buffer is used to reconstitute all other components.

The other components (except Separating reagent): Carefully add the required volume of assay buffer and replace stopper. Mix the contents of the bottles by inversion and swirling, taking care to avoid foaming.

Preparation of working standards

1. Label 6 polystyrene or polypropylene tubes 0.16, 0.31, 0.62, 1.25, 2.5 and 5.0.

2. Pipette 500 μl of assay buffer into all tubes.

3. Into the 5.0 tube pipette 500 μl of stock standard (100 ng/ml) and vortex thoroughly.

4. Transfer 500 μl from the 5.0 tube to the 2.5 tube and vortex thoroughly.

5. Repeat this doubling dilution successively with the remaining tubes.

6. 100 μl aliquots from each serial dilution give rise to 6 standard levels of rGH ranging from 0.16 ng to 5.0 ng/tube.

Note: One hundred microlitres (100 μl) of the reconstituted stock standard provided serves as the top standard (10 ng/tube). Working standards should be freshly prepared before each assay, and not re-used.

Assay procedure*(For a quick guide, refer to Table 1.)*

1. Equilibrate all reagents to room temperature.
2. Prepare reagents and assay standards as described in the previous section.
3. Label 12x75 mm disposable tubes in duplicate for total count (TC), non-specific binding (NSB), zero standard (Bo), standards and samples.
4. Pipette **200 μl** assay buffer into NSB tubes and **100 μl** assay buffer into Bo tubes.

5. Starting with the most dilute, pipette **100 µl** of each standard (S1-7) into the appropriately labelled tubes.
6. Pipette **100 µl** unknown sample (M_x) directly into appropriately labelled tubes.
7. Pipette **100 µl** antiserum into all tubes except NSB and TC.
8. Pipette **100 µl** of tracer into all tubes. The TC tubes should be stoppered and put aside for counting.
9. Vortex mix all tubes thoroughly. Cover the tubes, and incubate overnight (16-24 hours) at room temperature (15-30 °C).
10. Gently shake and swirl the bottle containing separating second antibody reagent (blue-green) to ensure a homogeneous suspension. Add **400 µl** into each tube except the TC. Vortex mix all tubes thoroughly and incubate at room temperature for 10 minutes.
11. Separate the antibody bound fraction using either magnetic separation or centrifugation as described below.

Magnetic separation

Attach the rack on the magnetic separator base and ensure that all tubes are in contact with the base plate. Leave for 15 minutes. After separation, do not remove the rack from the separator base. Pour off and discard the supernatant. Keeping the separator inverted, place the tubes on a pad of absorbent tissues and allow to drain for 5 minutes.

Centrifugation

Centrifuge all tubes at 4°C for 10 minutes at 1500 xg or greater. After centrifugation, place the tubes carefully into suitable decantation racks, then pour off and discard the supernatant. Keeping the tubes inverted, place the tubes on a pad of absorbent tissues and allow to drain for 5 minutes.

12. On completion of either magnetic or centrifugal separation, firmly blot the rims of the inverted tubes on the tissue pad to remove any adhering liquid. Do not re-invert the tubes once they have been turned upright.
13. Determine the radioactivity present in each tube by counting for at least 60 seconds in a gamma scintillation counter.

Table 1. Assay Protocol, Pipetting Guide (all volumes are in microlitres)

Tubes	TC	NSB	Bo	Stan - dard	Sam- ple
Buffer	-	200	100	-	-
Stan- dard	-	-	-	100	-
Sample	-	-	-	-	100
Anti- serum	-	-	100	100	100
Tracer	100	100	100	100	100
Vortex mix and incubate for 16-24 hours at room temperature (15-30 °C).					
Separating reagent	-	400	400	400	400
Vortex mix. Incubate for 10 minutes at room temperature.					
Separate either using magnetic separator for 15 minutes or by centrifugation for 10 minutes at >1500 xg .					
Decant tubes and blot on filter paper.					
Count radioactivity (60 sec/tube).					
Calculate the results.					

Calculation of results

Calculate the average count per minute (CPM) for each pair of assay tubes.

Calculate the percent NSB/TC using the following equation:

$$\text{NSB/TC(\%)} = \frac{\text{NSB (cpm)}}{\text{TC (cpm)}} \times 100$$

If the counter background is high, it should be subtracted from all counts.

Users may wish to subtract the average NSB cpm from all tubes except TC. If so the appropriate correction should be made. Calculate the percent Bo/TC using the following equation:

$$\text{Bo/TC(\%)} = \frac{\text{Bo (cpm)} - \text{NSB (cpm)}}{\text{TC (cpm)}} \times 100$$

Calculate the percent bound for each standard and sample using the following equation:

$$\text{B/Bo(\%)} = \frac{\text{S1-7/M}_x \text{ (cpm)} - \text{NSB (cpm)}}{\text{Bo (cpm)} - \text{NSB (cpm)}} \times 100$$

A standard curve can be generated by plotting the percent B/Bo as a function of the log rGH concentration.

Plot B/Bo(%) (y-axis) against concentration ng standard per tube (x-axis). The concentration (ng per tube) value of the samples can be read directly from the graph (see Figure 1).

Table 2. Typical assay data

Tube	Conc. ng/tube	Mean counts (cpm)	B/TC (%)	B/Bo (%)
TC	-	22837	-	-
NSB	-	927	4.1	-
Bo	-	12101	48.9	-
S1	0.16	11703	-	96.4
S2	0.31	11153	-	91.5
S3	0.62	9698	-	78.5
S4	1.25	7796	-	61.5
S5	2.5	5517	-	41.1
S6	5	3755	-	25.3
S7	10	2709	-	15.9

Note: The counts were obtained using new tracer. The counts will decline in line with the age of the tracer.

A decrease in Bo values will be observed during the shelf-life of this product. However, this drop in binding does not in our experience affect the curve parameters.

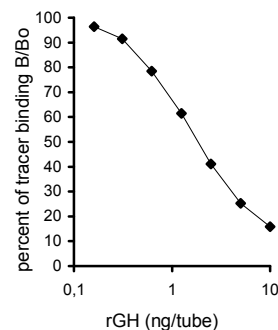


Figure 1: A typical standard curve (Do not use to calculate unknown samples!)

Characterization of assay

Stability

The components of this assay system will have a shelf-life of at least 2 weeks from the date of despatch.

Upon arrival, all components should be stored at 2-8°C where they are stable until the expiry date printed on the pack label. Once reconstituted, all reagents should be stored at 2-8 °C where they are stable for at least 14 days.

Non-specific binding

The non-specific binding (NSB) defined as the proportion of tracer bound in the absence of antibody was determined to be <5%.

The NSB was independent of tracer batch and did not change over a 10-week storage period.

Sensitivity

The sensitivity, defined as the amount of rGH needed to reduce zero dose binding by two standard deviations was 0.16 ng/tube.

Specificity

The cross-reactivity, as determined by the concentration giving 50% B/Bo with a number of related compounds is shown below.

Compound	Cross-reactivity (%)
rat GH	100
rat GH (NIDDK RP-2)	183
rat FSH (NIDDK RP-2)	0.042
rat LH (NIDDK RP-3)	0.13
rat PRL (NIDDK RP-3)	0.74
rat TSH (NIDDK RP-2)	0.15

Reproducibility

The intra-assay precision for duplicate determinations was calculated by repeatedly measuring an unknown in the assay. The result obtained was 1.32 ± 0.04 ng, CV = 3.0%. The between assay precision was assessed by repeated measurement of the same samples in successive assays using different batches of reagents and different operators. These results are shown below:

Sample	Number of replicates	Mean \pm 2SD	CV (%)
1	14	0.47 \pm 0.06	12.8
2	14	1.33 \pm 0.11	8.3
3	14	2.48 \pm 0.26	10.5

Dilutions of serum gave good parallelism with the standard curve (results from 2 experiments).

Sample volume (μ l)	serum	
	Measured conc. (ng/tube)	Calculated conc. (ng/ml)
25	0.541	21.6
50	0.88	17.78
100	1.65	16.50

Good recovery was obtained when rGH was added to plasma and serum:

Added conc. (ng / tube)	serum		
	Expected conc. (ng/tube)	Measured conc. (ng/tube)	Recovery (%)
0	-	1.65	-
0.95	2.60	2.37	91
1.82	3.47	3.02	87
3.33	4.98	4.45	89

Additional information

Components from various lots or from kits of different manufacturers should not be mixed or interchanged.

Precautions

Radioactivity

This product contains radioactive material. It is the responsibility of the user to ensure that local regulations or code of practice related to the handling of radioactive materials are satisfied.

Chemical hazard

Components contain sodium azide as an antimicrobial agent. Dispose of waste by flushing with copious amount of water to avoid build-up of explosive metallic azides in copper and lead plumbing. The total azide present in each pack is 158 mg.

All chemicals should be considered as potentially hazardous. We therefore recommend that this product is handled only by those persons who have been trained in laboratory techniques and that it is used in accordance with the principles of good laboratory practice. Wear suitable protective clothing such as laboratory overalls, safety glasses and gloves. Care should be taken to avoid contact with skin or eyes. In the case of contact with skin or eyes wash immediately with water (see safety data sheet for specific advice).

Safety data sheet

Product name:

Sodium azide

CAS No. 26628-22-8

R: 22-32 Toxic if swallowed. Contact with acids liberates very toxic gas.

S: (1/2)-28-45 (Keep locked up and out of the reach of children). After contact with skin, wash immediately with plenty of water. In case of accident or if you feel unwell, seek medical advice immediately (show label where possible).

Composition:

Sodium azide solution.

Hazards identification:

Toxic if swallowed, inhaled, or absorbed through skin. May cause eye and skin irritation.

First aid measures:

In case of contact, immediately flush eyes or skin with copious amounts of water. If inhaled remove to fresh air. In severe cases seek medical attention.

Fire fighting measures:

Dry chemical powder. Do not use water.

Accidental release:

Wear suitable protective clothing including laboratory overalls, safety glasses and gloves. Mop up spill area, place waste in a bag and hold for waste disposal. Wash spill site area after material pick-up is complete.

Handling and storage:

Wear suitable protective clothing including overalls, safety glasses and gloves. Do not get in eyes, on skin, or on clothing. Wash thoroughly after handling.

Personal protection:

See above instructions for handling and storage.

Physical and chemical properties:

Formula weight: 65.01. Density: 1.850.

Stability and reactivity:

Avoid contact with metals and acid chlorides. This yields a very toxic gas.

Toxicological information:

LD50: 27 mg/kg oral, rat

LD50: 20 mg/kg skin, rabbit

Ecological information:

Not applicable

Disposal consideration:

Up to 5 vials worth of material may be disposed of directly down the sink with water.

If 6 or more vials are to be disposed of they should pass through a chemical waste route.









Note: Inorganic azides will react with lead and copper plumbing fixtures to give explosive residues. Disposal of significant quantities of azides via such plumbing is not recommended.

Transport information :

No special considerations applicable.

Regulatory information:

The information contained in this safety data sheet is based on published sources and is believed to be correct. It should be used as a guide only. It is the responsibility of the user of this product to carry out an assessment of workplace risks, as may be required under national legislation.

	Use by	AS	Antiserum
	Batch code	CAL	Standard
	Caution, consult accompanying documents	SORB	Separating second antibody reagent
	Biological risk	TRAC	Tracer
	Consult operating instructions	BUF	Assay buffer
	Manufacturer		Temperature limitation Store between 2-8°C
REF	Catalogue number		Radioactive Material

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